

<p><b>Equipment Name:</b> <b>Auriga Focused Ion Beam</b></p>	<p><b>Category:</b></p> <p><b>B. Particle Labelling and/or</b> <b>C. Particle Characterisation in and ex-situ</b></p> <p><b>Institute: CRANN, Trinity College Dublin</b></p> <p><b>Location: CRANN Advanced Microscopy Laboratory, Unit 27-29, Trinity Enterprise Centre, Grand Canal Quay, Dublin 2, Ireland</b></p> <p><b>Contact Details of Technology Expert:</b>  <b>Name: Dr. Colm Faulkner</b>  <b>Phone: +353-1-896 4170</b>  <b>Fax,: +353-1-896 3037</b>  <b>E-mail: colm.faulkner@tcd.ie</b></p>
<p><b>Short technology description/Overview:</b></p> <p>The Carl Zeiss Auriga focused ion beam [FIB], is a one stop shop for nano-fabrication, 2-D and 3-D analysis and materials characterisation</p> <p>In a FIB, a beam of gallium ions, as narrow as 2.5 nm, is selectively written over a sample, acting as an ‘atomic chisel’ or ‘atomic power hose.’ The ion beam selectively etches away material, performing a rapid, dry, site specific, mask-less lithography with sub 5 nm patterning resolution. The system is “dual-beam” – a FIB to cut and a FESEM to image. The tool is perfect for nano and micro-fabrication, for example patterning prototype devices for micro-fluidics, or texturing a substrate for cell adhesion and movement investigations.</p> <p>The FIB is ideally suited to cross-sectioning material to resolve sub surface information, such as buried nano-particles, and has specialised software for acquiring 3-D information about samples using an automated slice and image function.</p> <p>The system has an in-situ micromanipulator, for placing/manipulating objects with sub 50 nm precision in-situ, and 5 different materials including Pt, W, and SiO<sub>2</sub> can be locally deposited in-situ, with a gas injections system [GIS]. Electron beam and ion beam lithography can be performed over large areas with Raith software package.</p> <p>The field emission electron column is capable of 1 nm imaging resolution at 30 KeV. It can operate down to 0.1 KeV – ideal for imaging challenging biological specimens and uncoated cells. Further by varying the energy of the electron beam intercellular structure, and embedded nano-particles in various samples can be imaged in a non-destructive manner.</p>	
<p><b>Main Features (Equipment Capabilities):</b></p> <ul style="list-style-type: none"> <li>• <b>3-D site specific reconstructions</b> – i.e. 3-D reconstruction of buried &amp; embedded nano-particles</li> <li>• <b>Site specific TEM sample preparation</b> – <i>in situ</i> and low KV capability to analyse buried nano-particles</li> <li>• <b>Site specific FIB cross-sections</b> – characterisation of embedded nano-particles and objects embedded in coatings, cells, and bio-materials, by site selective sputtering and FESEM imaging.</li> <li>• <b>Atom probe sample fabrication.</b> Fabrication of high aspect ratio pillars or material [potentially with embedded nano-particles], for high resolution characterisation by atom probe microscope or transmission electron microscope tomography.</li> <li>• <b>High Resolution Imaging:</b> Field emission scanning electron microscope [Spec – 1 nm, 0.3 KV- 30 KV]]</li> </ul>	

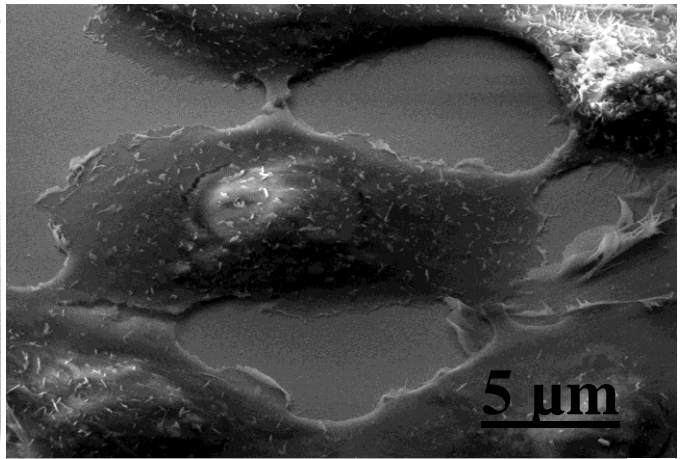
- **Cut:** 2.5 nm diameter gallium focused ion beam probe [FWHM].
- **Deposit:** Localized metal or insulator deposition, e.g. Pt, SiO<sub>2</sub>, W
- **Manipulate:** *In Situ* Kliendiek Micromanipulator
- **Pattern:** Elphi Quantum Lithography package for ion beam lithography or electron beam lithography

### Typical Samples & Images:

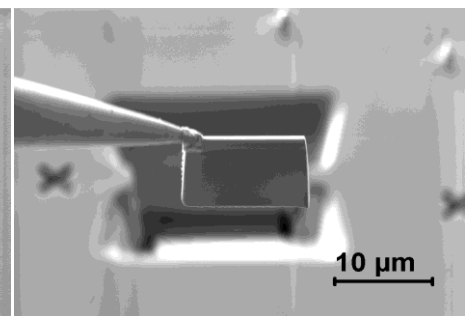
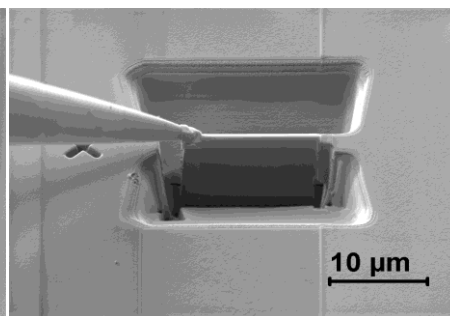
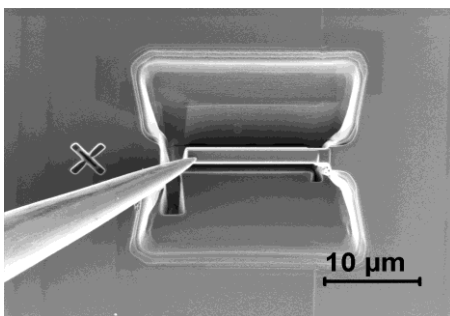
Some typical biological samples include uncoated and coated cells, collagen, drug impregnated polymers, elastin, candidate bone substitute materials, hydroxyapatite, platelets. Nano particles, composites, nanowires, nanotubes, and buried nano-particles for use in anti-bacterial coatings are all characterised. A wide range of silicon based samples are prepared for TEM.



FIB cross section of buried silver nano-particles in a polymer host material. [A. Satti, Y.K. Gun'ko, TCD]



FESEM imaging of uncoated cells. [D. Bazou, H. Z. Zhang, TCD]



FIB sample preparation on the TCD CRANN Auriga FIB.